micafungin MIC one to twofold dilutions above the CB (0.032, 0.064) were categorized as borderline resistant towards micafungin. Hence, 115 of these isolates were screened for mutations in *FKS1 HS1* and *HS2* regions utilizing next generation sequencing technology.

Results: In all 115 isolates, no missense mutations could be detected in *FKS1 HS1 or HS2*.

Conclusion: The micafungin resistance rate of *C. albicans* was considerably higher than in other echinocandins, although cross resistance is usually observed in echinocandin resistant isolates. No mutations were found in the hotspot regions of *FKS1* in these isolates indicating that resistance is very unlikely. Hence, it could be beneficial to revaluate the epidemiological cut-off value for micafungin for *C. albicans* and to reconsider the therapeutic failure evaluation for these isolates. Additionally, *in-vivo* studies should be done in order to evaluate the potential clinical failure.

S10-2 | Multiple versus single colony MIC testing in Candida species

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Background: Candida species is by far the most medical important fungal pathogen with antifungal drug resistance being on rise. Antifungal susceptibility testing (AFST) is recommended in patients who are unresponsive to treatment. For Aspergillus species it is advised to test at least 5 multiple colonies per plate, to secure strain-diversity in MICs to be analyzed.

Goal: The aim of this study was to compare single versus multiple colony MIC AFST, to check for MIC variability being present in macroscopically pure colonies.

Methods: In total, 100 yeast positive blood cultures (n = 56) and intra-abdominal fluids (n = 44) were investigated; AFST testing was done via E-test® applying fluconazole and anidulafungin. *Candida albicans* (n = 51), *C. glabrata* (n = 28), *C. dubliniensis* (n = 12), *C. tropicalis* (n = 8), *C. krusei* (n = 7), *C. parapsilosis* (n = 3), Saprochaete clavata (n = 2) and Cyberlindnera fabianii (n = 1) were studied.

Results: 500 MICs were obtained from 100 yeast strains, and only 3% showed MIC deviations. When testing 5 macroscopically identical colonies per plate, two *C. glabrata* strains displayed MIC inhomogeneity; MIC results highlighted fluconazole susceptible and resistant strains growing on one plate; another *C. albicans* strain harbored anidulafungin susceptible and resistant colonies.

Conclusion: In this set of yeasts investigated MIC inhomogeneity was low; AFST of multiple colonies seems to be not necessary in routine with the exception of special indications such treatment failure.

S10-3 | Disinfection of surfaces contaminated with *Candida auris* using ozone under complex room conditions

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Introduction: *Candida auris* is an emerging multiresistant yeast, which has caused multiple hospital outbreaks. Yeasts can display a high ability to survive on inanimate surfaces. Therefore, cleaning and disinfection is an important part in the prevention of *C. auris* transmission. In this study, we investigated the efficacy of an automated room decontamination using ozone against *C. auris* under complex room conditions.

Material/Methods: Four *C. auris* strains (10111031/south-asian clade I, 10111016/Latin American clade IV, 10111005/Austria, 10110211/Netherlands) were analyzed with respect of their ability to survive on dry surfaces. Two strains with a high survival rate were used for further experiments. The Sterisafe™ pro instrument was used in a patient room with an attached bathroom. *C. auris* was dried on three different carriers (ceramic tiles, stainless steel, solid core furniture board) and placed at eight different positions in the rooms. A standard disinfection cycle (80 ppm ozone; 90% RH; 60 min) was conducted in three independent experiments. *Candida albicans* ATCC® 10231™ recommended to confirm yeasticidal activity was used as control.

Results: *C. auris* 10111005 rapidly died off during desiccation on all surfaces. The other strains displayed different killing rates during initial desiccation but cell counts remained stable on the surfaces for one day. Strains 10111031 and 10111016 reached sufficient cell counts for further disinfection experiments. For all three yeasts investigated, mean reduction rates greater than 4 log were reached even under the complex room conditions using the standard disinfection cycle of the SterisafeTM pro instrument.

Conclusions: *C. auris* strains display a differential behavior on dry inanimate surfaces with three strains displaying high survival rates. Ozone generated with the Sterisafe™ pro instrument displayed a sufficient yeasticidal activity against the tested *C. auris* strains as well as against *C. albicans*.